

Omnia E Hussein¹, Mousa O Germoush² and Ayman M Mahmoud^{1*}

¹Physiology Division, Zoology Department, Faculty of Science, Beni-Suef University, Beni-Suef, Egypt

²Department of Biology, Faculty of Science, Aljouf University, Sakaka, Saudi Arabia

Dates: Received: 22 December, 2015; Accepted: 20 February, 2016; Published: 22 February, 2016

*Corresponding author: Ayman M. Mahmoud, PhD Physiology Division, Zoology Department, Faculty of Science, Beni-Suef University, Salah Salim St, 62514, Beni-Suef, Egypt. Tel.: +201144168280; E-mail: aymano911@yahoo.com; ayman.mahmoud@science.bsu.edu.eg

www.peertechz.com

Keywords: Renal injury; Rue; Oxidative stress; Inflammation; Anti-tuberculosis therapy

Research Article

Ruta graveolens Protects Against Isoniazid/Rifampicin-Induced Nephrotoxicity Through Modulation of Oxidative Stress and Inflammation

Abstract

Background and Aim: Drug-induced nephrotoxicity is a renal dysfunction that arises as a result of exposure to nephrotoxic drugs. Anti-tuberculosis therapy can cause nephrotoxicity and permanent kidney damage. The current study was designed to evaluate the possible protective effects of *Ruta graveolens* L. leaves extract against isoniazid/rifampicin-induced nephrotoxicity in rats.

Methods: The experimental rats received isoniazid and rifampicin at dose level of 50 mg/kg, and 50 or 100 mg/kg/day *Ruta graveolens* leaves extract orally for 45 days.

Results: Isoniazid/rifampicin administration induced kidney injury evidenced by the histopathological alterations as well as significant ($P < 0.001$) increase in serum creatinine, urea and uric acid. Isoniazid/rifampicin-intoxicated rats exhibited a significant increase in serum tumor necrosis factor alpha ($P < 0.001$), and renal lipid peroxidation ($P < 0.01$) and nitric oxide ($P < 0.001$) levels. On the other hand, reduced glutathione level, and activity of superoxide dismutase and glutathione peroxidase were markedly ($P < 0.001$) declined in kidney of isoniazid/rifampicin-induced rats. Concomitant supplementation with either 50 or 100 mg/kg dose of *Ruta graveolens* leaves extract prevented the isoniazid/rifampicin-induced biochemical and histopathological alterations.

Conclusion: The present investigation confers new information on the protective mechanism of *Ruta graveolens* leaves extract on anti-tuberculosis therapy-induced nephrotoxicity. *Ruta graveolens* protects against isoniazid/rifampicin-induced renal injury through attenuation of inflammation and oxidative stress, and potentiation of the antioxidant defense system.

Introduction

The kidney is a vital organ required to essential functions including regulation of the extracellular environment, maintenance of homeostasis, and detoxification and excretion of drugs and toxic metabolites [1]. The kidneys are therefore vulnerable to drug-induced toxicity. Drug-induced nephrotoxicity is a renal dysfunction that arises as a direct or indirect result of exposure to drugs [2]. Nephrotoxic drugs are therapeutic agents that have the potential to induce adverse effects on renal function due to direct toxicity or compromised renal perfusion [3]. Previous studies showed that nephrotoxic drugs were responsible for 19%–25% of acute kidney injury in critically ill patients [4–6]. Nephrotoxic drug-induced kidney dysfunction include acute tubular necrosis, haemodynamically mediated damage, glomerular and tubulointerstitial injury and obstructive nephropathy [7].

The use of anti-tuberculosis drugs, isoniazid and rifampicin, has been reported to be associated with hepatotoxicity that could result in acute liver injury and a high mortality rate [8,9]. The rate of their hepatotoxic effects is much higher in developing countries than that in developed countries [10]. We have demonstrated the contribution of oxidative stress and inflammation in isoniazid-induced hepatotoxicity [9]. In addition, studies have reported that reactive oxygen species and oxidative stress play a key role in the pathogenesis

of drug-induced renal damage [11,12]. Although hepatotoxicity of the anti-tuberculosis drugs has been extensively studied, their induced nephrotoxicity has been poorly documented. Recently, we reported that isoniazid and rifampicin combination induced nephrotoxicity associated with oxidative stress and inflammation in rats [13].

Medicinal plants are broadly used in the treatment of several diseases owing their cheapness, safety and nontoxicity when compared with the synthetic drugs [14]. *Ruta graveolens* L. (Family: *Rutaceae*) is an ancient medicinal plant. It is commonly known as rue or sadab and currently used for treatment of eye problems, dermatitis, aching pain, rheumatism, psoriasis, multiple sclerosis, leucoderma and cutaneous lymphomas [15]. Multiple studies have reported that rue possesses hepatoprotective, anticancer, anti-inflammatory, antioxidant, anti-hyperammonemic and antidiabetic activities [16–19]. Recently, we demonstrated the protective effect of *R. graveolens* leaves extract against diethylnitrosamine-induced kidney damage in rats [20]. Although the beneficial effects of *R. graveolens* in multiple disease cases have been reported, its protective effect against nephrotoxicity associated with the use of anti-tuberculosis drugs has not been studied. Therefore, the present study was designed to scrutinize the protective effects of *R. graveolens* leaves extract against isoniazid/rifampicin-induced renal injury in rats, focusing on oxidative stress and inflammation.

Materials and Methods

Chemicals

Isoniazid (INH) was supplied by El Nasr Company for Chemicals and Drugs (Cairo, Egypt), and rifampicin (RIF) was purchased from Novartis Pharma Company (Cairo, Egypt). Reduced glutathione (GSH), pyrogallol, trichloroacetic acid (TCA), thiobarbituric acid (TBA) and 5,5'-dithiobis-(2-nitrobenzoic acid) (DTNB) were purchased from Sigma (USA). All other chemicals were of analytical grade and obtained from standard commercial supplies.

Collection of plant and extract preparation

R. graveolens (sadab) was collected and the extract was prepared as we reported previously [19]. Briefly, sadab was obtained from the Experimental Station of Medical Plants (ESMP), Faculty of Pharmacy, Cairo University (Egypt). The plant leaves were collected, cleaned, air dried and ground with an electric grinder. The powdered leaves were macerated in 80% aqueous ethanol for 24 h at room temperature. Following filtration, the filtrate was concentrated under vacuum in a rotary evaporator. The residue obtained was stored frozen till use.

Animals and treatments

Male Wistar rats weighing 140-160 g, obtained from animal house of the National Research Centre (El-Giza, Egypt) were included in the present investigation. The animals were housed in plastic well-aerated cages (6 rats/cage) at normal atmospheric temperature ($25 \pm 2^\circ\text{C}$) and normal 12 h light/dark cycle. Rats had free access to water and were supplied daily with laboratory standard diet of known composition *ad libitum*. All animal procedures were undertaken with the approval of Institutional Animal Ethics Committee of Beni-Suef University (Egypt).

Twenty-four rats were divided to four groups (N = 6) and were subjected to the following treatments:

Group I (Control): Rats received the vehicle 0.5% carboxymethylcellulose (CMC) via oral gavage for 45 days and served as control.

Group II (INH/RIF): Rats received isoniazid (50 mg/kg body weight), rifampicin (50 mg/kg body weight) and 0.5% CMC orally for 45 days [13].

Group III (INH/RIF + 50 mg *R. graveolens*): Rats received isoniazid (50 mg/kg), rifampicin (50 mg/kg) and 50 mg/kg body weight *R. graveolens* leaves extract dissolved in 0.5% CMC orally for 45 days [20].

Group IV (INH/RIF + 100 mg *R. graveolens*): Rats received isoniazid (50 mg/kg) and rifampicin (50 mg/kg) dissolved in water and 100 mg/kg body weight *R. graveolens* leaves extract dissolved in 0.5% CMC orally for 45 days [19].

The doses of INH, RIF and *R. graveolens* were balanced consistently over the entire period of study, as indicated by any change in body weight.

Samples preparation

By the end of the experiment, rats were sacrificed under ether anesthesia and blood samples were collected, left to coagulate and centrifuged at 3000 rpm for 15 min to separate serum. Kidneys were immediately excised, perfused with ice-cold saline, and samples were kept frozen or fixed in 10% buffered formalin for histological processing. Frozen samples (10% w/v) were homogenized in chilled phosphate buffered saline and the homogenates were centrifuged at 3000 rpm for 10 min. The clear homogenates were collected and used for assaying oxidative stress and antioxidant defenses.

Biochemical assays

Determination of serum creatinine, urea and uric acid: Serum creatinine, urea and uric acid levels were assayed using reagent kits purchased from Biosystems (Spain), following the methods of Young [21], Kaplan [22] and Fossati et al. [23], respectively.

Determination of tumor necrosis factor-alpha (TNF- α): Serum levels of TNF- α were determined by specific ELISA kits purchased from R&D Systems (USA), according to the manufacturer's instructions. The concentration of TNF- α was determined spectrophotometrically at 450 nm. Standard plot was constructed by using standard cytokine and the concentration for unknown samples was calculated from the standard plot.

Determination of oxidative stress and antioxidant defenses: Lipid peroxidation, assayed as malondialdehyde (MDA), was determined in kidney homogenate according to the method of Preuss et al. [24]. Nitric oxide level was estimated in the kidney homogenate as nitrite according to the method of Montgomery and Dymock [25], using reagent kit purchased from Biodiagnostics (Egypt). The assay is based on the Griess method which is a simple colorimetric reaction between nitrite, N-(1-naphthyl) ethylenediamine and sulfonamide to produce a pink product with maximum absorbance at 543 nm. Reduced glutathione (GSH) content was assayed according to the method of Beutler et al. [26]. Glutathione peroxidase (GPx) and superoxide dismutase (SOD) activities were measured according to the methods of Matkovic et al. [27] and Marklund and Marklund [28], respectively.

Histopathological study

Kidney samples were flushed with cold saline and then fixed in 10% buffered formalin for at least 24 h. The specimens were then dehydrated in ascending series of ethanol, cleared in xylene and embedded in paraffin wax. Blocks were prepared and 4 μm thick sections were cut by a sledge microtome. The paraffin embedded sections were deparaffinized, washed and stained with hematoxylin and eosin (H&E). The stained slides were examined under light microscope.

Statistical analysis

Statistical analysis was performed using GraphPad Prism 5 software (GraphPad Software, San Diego, CA, USA). Results were expressed as mean \pm standard deviation (SD) and all statistical comparisons were made by means of the one-way ANOVA test followed by Tukey's test *post hoc* analysis. A P value <0.05 was considered significant.

Table 1: Serum creatinine, urea and uric acid levels in control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*.

	Creatinine (mg/dl)	Urea (mg/dl)	Uric acid (mg/dl)
Control	0.87 ± 0.04	23.90 ± 3.78	1.54 ± 0.30
INH/RIF	1.56 ± 0.22***	65.93 ± 2.59***	4.21 ± 0.99***
INH/RIF + 50 mg <i>R. graveolens</i>	0.93 ± 0.13###	30.21 ± 2.72###	2.15 ± 0.61##
INH/RIF + 100 mg <i>R. graveolens</i>	0.90 ± 0.13###	31.34 ± 4.12###	1.63 ± 0.22###

Data are expressed as M ± SD. ***P<0.001 vs control, and ###P<0.01 and ####P<0.001 vs INH/RIF. INH, isoniazid; RIF, rifampicin; *R. graveolens*, *Ruta graveolens*; SD, standard deviation; vs, versus.

Results

Data summarized in Table 1 show the effect of isoniazid/rifampicin administration and treatment with *R. graveolens* leaves extract on renal function markers. The administration of isoniazid/rifampicin produced marked impairment of kidney function as showed by the significant (P<0.001) increase in serum creatinine, urea and uric acid levels. Concurrent oral administration of 50 mg/kg *R. graveolens* leaves extract significantly decreased the elevated levels of serum creatinine (P<0.001), urea (P<0.001) and uric acid (P<0.01) when compared with the isoniazid/rifampicin control group. Similarly, administration of 100 mg/kg *R. graveolens* significantly (P<0.001) alleviated serum levels of creatinine, urea and uric acid in isoniazid/rifampicin-induced rats.

Serum levels of TNF-α exhibited a significant (P<0.001) increase in isoniazid/rifampicin-intoxicated rats when compared with the control group, as depicted in Figure 1. Although non-significant (P>0.05), concomitant supplementation of the 50 mg/kg *R. graveolens* leaves extract decreased serum TNF-α. On the other hand, administration of the higher *R. graveolens* dose (100 mg/kg) produced a significant (P<0.001) decrease in circulating levels of TNF-α when compared with the isoniazid/rifampicin-administered rats.

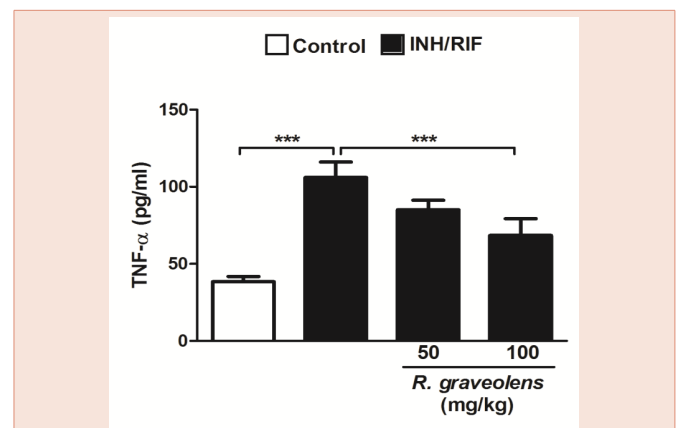
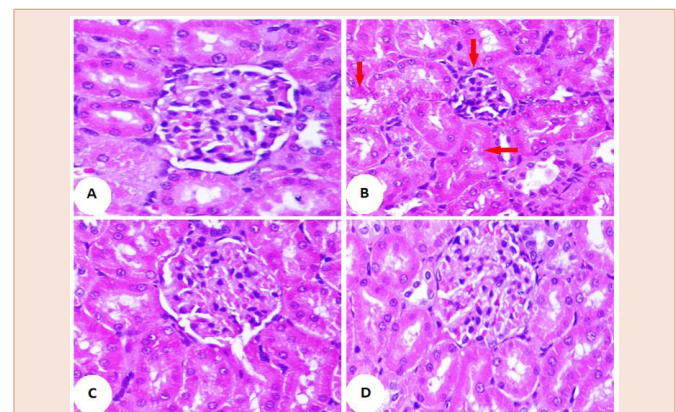
Histopathological examination of the kidney sections of control rats showed normal histological structure of the renal tissue (Figure 2A). Conversely, isoniazid/rifampicin-induced rats showed several histopathological changes and lesions, including atrophy of glomerular tuft, dysplastic renal tubules and inflammatory cells infiltration (Figure 2B). Concurrent treatment of the isoniazid/rifampicin-administered rats with either 50 or 100 mg/kg *R. graveolens* leaves extract produced marked improvement in the kidney histological structure and prevented the isoniazid/rifampicin-induced alterations, as represented in Figure 2C and 2D, respectively.

Concerning lipid peroxidation, rats received isoniazid/rifampicin orally for 45 days exhibited significantly (P<0.001) increased renal lipid peroxidation levels when compared with the normal control rats, as represented in Figure 3. Concurrent oral administration of *R. graveolens* at dose levels of both 50 and 100 mg/kg produced a significant (P<0.001) amelioration in the lipid peroxidation marker, MDA, in isoniazid/rifampicin-induced rats.

Renal nitric oxide exhibited a similar pattern. It showed a significant (P<0.001) increase in kidney of isoniazid/rifampicin-administered rats. Oral administration of either 50 or 100 mg/kg *R.*

graveolens leaves extract markedly (P<0.05) decreased renal nitric oxide levels in isoniazid/rifampicin-induced rats, as depicted in Figure 4.

On the contrary, 45 days of isoniazid/rifampicin administration significantly (P<0.001) decreased kidney GSH content when compared with the control group (Figure 5). Concurrent supplementation of either 50 or 100 mg/kg *R. graveolens* leaves extract produced a significant (P<0.001) alleviation of renal GSH content. Similarly, GPx activity showed a significant (P<0.001) decrease in the kidney of isoniazid/rifampicin-induced rats, as represented in Figure 6. SOD activity as well was significantly (P<0.001) decreased in the kidney of isoniazid/rifampicin-induced rats when compared with the control group (Figure 7). Oral supplementation of 50 mg/kg *R. graveolens* leaves extract potentially (P<0.01) ameliorated the activity of GPx and SOD in kidney of isoniazid/rifampicin-intoxicated rats. At the higher dose (100 mg/kg), *R. graveolens* leaves extract significantly enhanced the activity of GPx (P<0.001) and SOD (P<0.01) in kidney of isoniazid/rifampicin-induced rats.


Figure 1: Serum TNF-α levels in control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M ± SD. ***P<0.001. INH, isoniazid; RIF, rifampicin; TNF-α, tumor necrosis factor alpha; SD, standard deviation.

Figure 2: Photomicrographs of H&E stained kidney sections of control (A [x400]) showing normal histological structure, INH/RIF (B [x400]) showing dysplastic renal tubules, and atrophy of glomerular tuft, INH/RIF + 50 mg *Ruta graveolens* (C [x400]), and INH/RIF + 100 mg *Ruta graveolens* (D [x400]) showing nearly normal renal tubules and renal corpuscles.

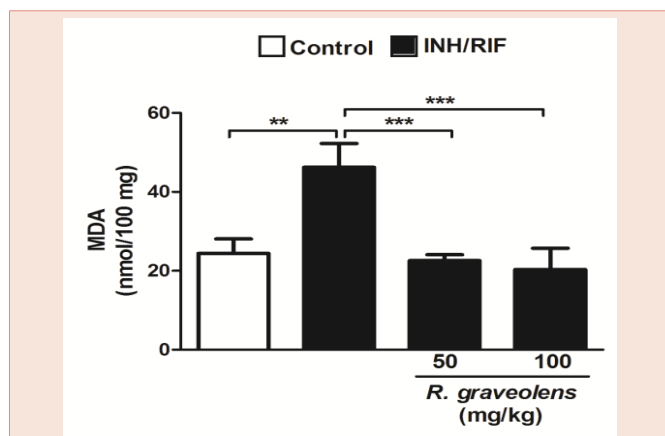


Figure 3: Lipid peroxidation levels in kidneys of control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M \pm SD. **P<0.01 and ***P<0.001. INH, isoniazid; RIF, rifampicin; MDA, malondialdehyde; SD, standard deviation.

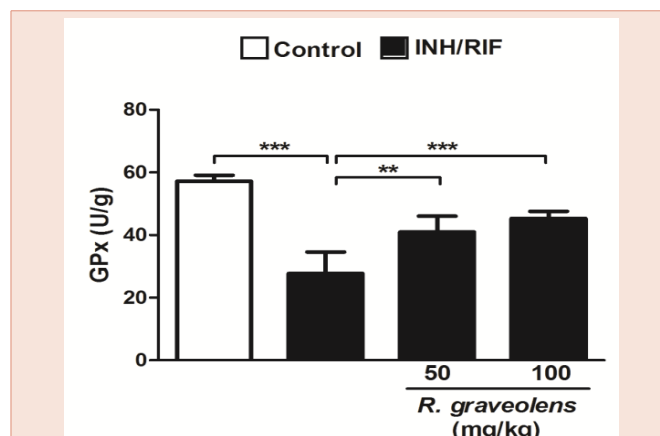


Figure 6: Glutathione peroxidase activity in kidneys of control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M \pm SD. ***P<0.001. INH, isoniazid; RIF, rifampicin; GPx, glutathione peroxidase; SD, standard deviation.

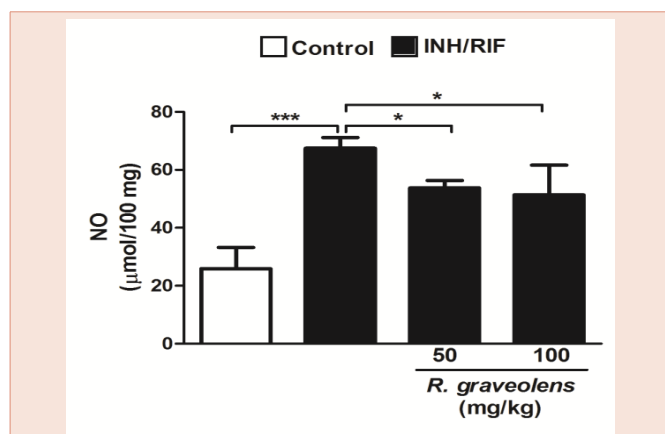


Figure 4: Nitric oxide levels in kidneys of control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M \pm SD. *P<0.05 and ***P<0.001. INH, isoniazid; RIF, rifampicin; NO, nitric oxide; SD, standard deviation.

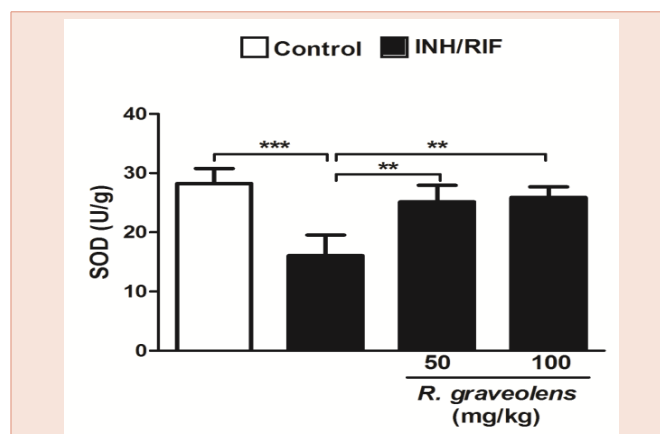


Figure 7: Superoxide dismutase activity in kidneys of control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M \pm SD. **P<0.01 and ***P<0.001. INH, isoniazid; RIF, rifampicin; SOD, superoxide dismutase; SD, standard deviation.

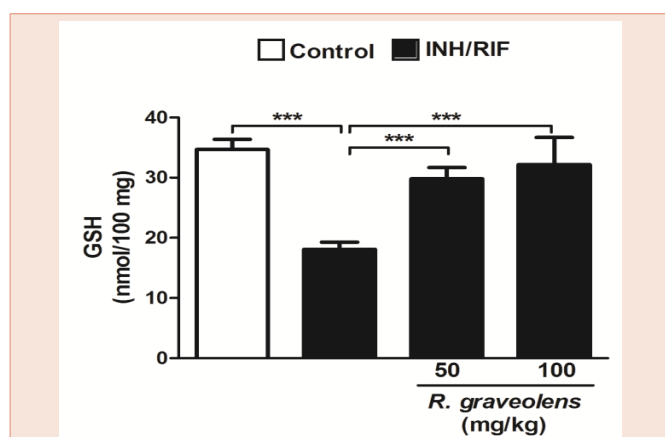


Figure 5: Reduced glutathione in kidneys of control, INH/RIF and INH/RIF rats treated with *Ruta graveolens*. Data are expressed as M \pm SD. ***P<0.001. INH, isoniazid; RIF, rifampicin; GSH, glutathione; SD, standard deviation.

Discussion

In this investigation, we evaluated the protective effects of *R. graveolens* leaves extract on isoniazid/rifampicin-induced nephrotoxicity in rats. We found for the first time that concurrent treatment with *R. graveolens* protected against kidney injury through attenuation of inflammation and oxidative stress, and potentiating the antioxidant defenses.

Nephrotoxicity and nephropathy are important microvascular complication of anti-tuberculosis therapy [9,13,29]. Multiple studies have demonstrated that both inflammation and oxidative stress are implicated in drug-induced organ injury [9,13,30]. Rekha et al. [29], reported that anti-tuberculosis therapy-induced renal injury is usually reversible and could be treated properly if detected early. In this context, Mahmoud et al. [13], suggested that the use of antioxidants could protect against isoniazid/rifampicin-induced oxidative stress and nephrotoxicity. Therefore, we have attempted to test the

hypothesis that *R. graveolens* leaves extract could protect against isoniazid/rifampicin-induced renal damage through attenuation of inflammation and prevention of oxidative stress.

The present findings showed that administration of isoniazid and rifampicin has induced kidney injury and glomerular dysfunction evidenced by the elevated circulating creatinine, urea and uric acid levels. These measurements are often regarded as reliable markers of kidney damage [31] and indicate the loss of a majority of kidney function [32]. These elevated assessments are in agreement with the studies of Hashmi et al. [33], who reported increased serum levels of creatinine and urea in albino rabbits following administration of anti-tuberculosis drugs. More recently, we demonstrated elevated serum creatinine, urea and uric acid in serum of isoniazid/rifampicin-induced rats [13]. Isoniazid/rifampicin-induced nephrotoxicity was further confirmed by the recorded histological alterations including atrophy of glomerular tuft, dysplastic renal tubules and inflammatory cells infiltration, as we recently reported [13]. Concomitant administration of either dose of *R. graveolens* leaves extract significantly decreased circulating creatinine, urea and uric acid levels, and potentially attenuated the induced histological alterations in the kidney. Accordingly, we have recently demonstrated that *R. graveolens* at dose level of 50 mg/kg markedly decreased circulating kidney function markers and prevented histopathological alterations in diethylnitrosamine-induced nephrotoxicity in rats [20]. We have also demonstrated that at dose level of 100 mg/kg, *R. graveolens* was able to decrease serum urea levels in ammonium chloride-induced hyperammonemic rats [19].

Isoniazid and rifampicin administration induced a significant increase in serum levels of the pro-inflammatory cytokine TNF- α which represents an important mediator of inflammatory tissue damage. Studies presented evidence that nephrotoxicants could provoke an inflammatory response leading to organ injury [12,34]. The significantly elevated TNF- α reflects the degree of inflammation. In this context, our previous studies demonstrated a marked increase in serum pro-inflammatory cytokines in anti-tuberculosis drugs administered rats [9,13]. In a dose-dependent manner, concurrent administration of *R. graveolens* leaves extract produced pronounced decline in serum TNF- α , indicating its anti-inflammatory efficacy. These findings are in agreement with our recent study where we reported the ability of *R. graveolens* to decrease circulating levels of TNF- α in hyperammonemic rats [19].

The kidneys are vulnerable to damage caused by reactive oxygen species due to the abundant long chain polyunsaturated fatty acids in the composition of their membrane lipids [35]. Here, isoniazid/rifampicin-induced rats showed marked elevation of the lipid peroxidation marker, MDA, and nitric oxide, revealing oxidative as well as nitrosative stress. Under oxidative stress conditions, reactive oxygen species induce DNA fragmentation, cellular injury, protein damage and lipid peroxidation [36,37]. In addition, isoniazid/rifampicin-induced rats exhibited significantly declined renal GSH content which is a potent antioxidant forming S-conjugates with products of lipid peroxidation [38]. Furthermore, activity of the antioxidant enzymes SOD and GPx was declined in kidneys of the isoniazid/rifampicin-induced rats. These enzymes play an important

role in protecting the body against the deleterious effects of reactive oxygen species [39]. Accordingly, we have demonstrated increased lipid peroxidation and declined antioxidant defenses in kidney of isoniazid/rifampicin-administered rats [13]. Concomitant treatment with *R. graveolens* leaves extract markedly decreased kidney lipid peroxidation and nitric oxide, and enhanced the antioxidant defenses, suggesting a potent free radical scavenging and antioxidant efficacies. We have previously showed the antioxidant and renoprotective effects of *R. graveolens* in hyperammonemic and diethylnitrosamine-induced rats [19,20]. Therefore, we assume that the protective mechanism of *R. graveolens* against isoniazid/rifampicin-induced nephrotoxicity is mediated partially through prevention of GSH decline and potentiation of the antioxidant enzymes.

In conclusion, the present investigation confers new information on the protective mechanism of *R. graveolens* leaves extract against anti-tuberculosis drug-induced nephrotoxicity. This renoprotective effect could be attributed to the ability of *R. graveolens* to attenuate inflammation, inhibit lipid peroxidation, prevent GSH decline and enhance the enzymatic antioxidant defenses.

References

- Ferguson MA, Vaidya VS, Bonventre JV (2008) Biomarkers of nephrotoxic acute kidney injury. *Toxicology* 245: 182-193.
- Finn W, Porter G (2003) *Urinary biomarkers and nephrotoxicity*. Clinical Nephrotoxins, 2nd ed. Kluwer Academic Publishers, Massachusetts 621-655.
- Finlay S, Bray B, Lewington AJ, Hunter-Rowe CT, Banerjee A, et al. (2013) Identification of risk factors associated with acute kidney injury in patients admitted to acute medical units. *Clin Med* 13: 233-238.
- Mehta RL, Pascual MT, Soroko S, Savage BR, Himmelfarb J, et al. (2004) Spectrum of acute renal failure in the intensive care unit: The PICARD experience. *Kidney Int* 66: 1613-1621.
- Uchino S, Kellum JA, Bellomo R, Doig GS, Morimatsu H, et al. (2005) Acute renal failure in critically ill patients: A multinational, multicenter study. *JAMA* 294: 813-818.
- Naughton CA (2008) Drug-induced nephrotoxicity. *Am Fam Phys* 78: 743-750.
- Taber SS, Mueller BA (2006) Drug-associated renal dysfunction. *Crit Care Clin* 22: 357-374.
- Kumar R, Shalimar, Bhatia V, Khanal S, Sreenivas V, et al. (2010) Antituberculosis therapy-induced acute liver failure: magnitude, profile, prognosis, and predictors of outcome. *Hepatology* 51: 1665-1674.
- Mahmoud A M, Germoush MO, Soliman AS (2014) Berberine attenuates isoniazid-induced hepatotoxicity by modulating peroxisome proliferator-activated receptor γ , oxidative stress and inflammation. *Int J Pharmacol* 10: 451-460.
- Pal R, Vaiphei K, Sikander A, Singh K, Rana SV. (2006) Effect of garlic on isoniazid and rifampicin induced hepatic injury in rats. *World J Gastroenterol* 12: 636-639.
- Lopez-Novoa JM, Quiros Y, Vicente L, Morales AI, Lopez-Hernandez FJ (2011) New insights into the mechanism of aminoglycoside nephrotoxicity: an integrative point of view. *Kidney Int* 79: 33-45.
- Mahmoud AM, Galaly SR, Ahmed OM. (2014): Thymoquinone and curcumin attenuate gentamicin-induced renal oxidative stress, inflammation and apoptosis in rats. *EXCLI J* 13: 98-110.
- Mahmoud AM, Morsy BM, Abdel-Hady DS, Samy RM (2015) *Prunus armeniaca* leaves extract protects against isoniazid and rifampicin induced

- nephrotoxicity through modulation of oxidative stress and inflammation. *Int J Food Nutr Sci* 2: 1-6.
14. Bhawna S, Kumar SU (2009) Hepatoprotective activity of some indigenous plants. *Int J Pharm Tech* 1: 1330-1334.
 15. Miguel ES (2003) Rue in traditional Spain: frequency and distribution of its medicinal and symbolic applications. *Econ Bot* 57: 231-244.
 16. Pathak S, Multani A, Banerji P, Banerji P (2003) Ruta 6 selectively induces cell death in brain cancer cells but proliferation in normal peripheral blood lymphocytes: a novel treatment for human brain cancer. *Int J Oncol* 23: 975-982.
 17. Preethi K, Kuttan G, Kuttan R (2006) Anti-tumour activity of *Ruta graveolens* extract. *Asian Pac J Cancer Prev* 7: 439-443.
 18. Ahmed OM, Moneim AA, Yazid IA, Mahmoud AM (2010) Antihyperglycemic, antihyperlipidemic and antioxidant effects and the probable mechanisms of action of *Ruta graveolens* infusion and rutin in nicotinamide-streptozocin induced diabetic rats. *Diabetol Croat* 39: 15-35.
 19. Mahmoud AM, Germoush MO, Soliman AS (2014) *Ruta graveolens* mitigates ammonium chloride-induced hyperammonemia by modulating antioxidant status and pro-inflammatory cytokines. *Life Sci J* 11: 269-275.
 20. Mahmoud AM, Ahmed RR, Soliman HA, Salah M (2015) *Ruta graveolens* and its active constituent rutin protect against diethylnitrosamine-induced nephrotoxicity through modulation of oxidative stress. *J App Pharm Sci* 5: 16-21.
 21. Young DS (1995) Effects of drugs on Clinical Lab. Tests, 4th ed AACC Press.
 22. Kaplan A (1984) Urea. *Clin Chem The CV Mosby Co St Louis. Toronto. Princeton*, 1257-1260 and 437 and 418.
 23. Fossati P, Prencipe L, Berti G (1980) Use of 3,5-dichloro-2-hydroxybenzenesulfonic acid/4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. *Clin Chem* 26: 227-231.
 24. Preuss HG, Jarrell ST, Scheckenbach R, Lieberman S, Anderson RA (1998) Comparative effect of chromium vanadium and Gymnemasylvestre on sugar-induced blood pressure elevation in SHR. *J Am Coll Nutr* 17: 116-123.
 25. Montgomery HAC, Dymock JF (1961) The determination of nitrite in water. *Analyst* 86: 414-416.
 26. Beutler E, Duron O, Kelly BM (1963) Improved method for the determination of blood glutathione. *J Lab Clin Med* 61: 882-888.
 27. Matkovics B, Szabo L, Varga IS (1998) Determination of enzyme activities in lipid peroxidation and glutathione pathways (in Hungarian). *Laboratoriumi Diagnosztika* 15: 248-249.
 28. Marklund SL, Marklund G (1974) Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur J Biochem* 47: 469-474.
 29. Rekha VVB, Santha T, Jawahar MS. (2005) Rifampicin induced renal toxicity during retreatment of patients with pulmonary tuberculosis. *JAPI* 53: 811-813.
 30. Mahmoud AM (2014) Hesperidin protects against cyclophosphamide-induced hepatotoxicity by upregulation of PPAR γ and abrogation of oxidative stress and inflammation. *Can J Physiol Pharmacol* 92: 717-724.
 31. Adebisi SA, Oluboyo PO, Okesina AB (2000) Effect of drug-induced hyperuricaemia on renal function in Nigerians with pulmonary tuberculosis. *Afr J Med Med Sci* 3-4:297-300.
 32. Rached E, Hoffmann D, Blumbach K, Weber K, Dekant W, et al. (2008) Evaluation of putative biomarkers of nephrotoxicity after exposure to ochratoxin A in vivo and in vitro. *Toxicol Sci* 103: 371-381.
 33. Hashmi N, Muhammad F, Javed I, Khan JA, Khan MZ, et al. (2013) Nephroprotective effects of *Ficus religiosa* linn (peepal plant) stem bark against isoniazid and rifampicin induced nephrotoxicity in albino rabbits. *Pak Vet J* 33: 330-334.
 34. Araujo LP, Truzzi RR, Mendes GE, Luz MA, Burdmann EA, et al. (2012) Annexin A1 protein attenuates cyclosporine-induced renal hemodynamics changes and macrophage infiltration in rats. *Inflamm Res* 61: 189-196.
 35. Ozbek E (2012) Induction of oxidative stress in kidney. *Int J Nephrol* 2012: 465897.
 36. Mittal G, Brar AP, Soni G (2006) Impact of hypercholesterolemia on toxicity of Nnitrosodiethylamine: Biochemical and histopathological effects. *Pharmacol Rep* 58:413-419.
 37. Nencini C, Giorgi G, Micheli L (2007) Protective effect of silymarin on oxidative stress in rat brain. *Phytomedicine* 14: 129-135.
 38. Laurent A, Perdu-Durand E, Alary J, Debrauwer L, Cravedi JP (2000) Metabolism of 4-hydroxynonenal, a cytotoxic product of lipid peroxidation in rat precision-cut liver slices. *Toxicol Lett* 114: 203-214.
 39. Wei XJ, Hu TJ, Chen JR, Wei YY (2011) Inhibitory effect of carboxymethylpachymaran on cyclophosphamide-induced oxidative stress in mice. *Int J Biol Macromol* 49: 801-805.

Copyright: © 2016 Hussein OE, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Hussein OE, Germoush MO, Mahmoud AM (2016) *Ruta graveolens* Protects Against Isoniazid/Rifampicin-Induced Nephrotoxicity through Modulation of Oxidative Stress and Inflammation. *Glob J Biotechnol Biomater Sci* 2(1): 008-013.